## Histomorphometric Analysis of Skin and Stress Indices of Nile Tilapia (*Oreochromis niloticus*) Exposed to Different Concentrations of Ammonia

#### Masuomeh Rahmati<sup>1</sup>, Hassan Morovvati<sup>1\*</sup>, Rahim Abdi<sup>2</sup>

1. Department of Basic Sciences, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran

2. Department of Marine Biology, Faculty of Marine Science, Khorramshahr University of Marine Science and Technology, Khorramshahr, Iran

## Abstract

**BACKGROUND:** Intensive aquaculture, which is always associated with high concentrations of nitrogen pollutants, can lead to increased economic damage and energy loss, and increased fish mortality.

**OBJECTIVES:** This study aimed to investigate the histomorphometry of the skin and stress indices of the Nile tilapia on exposure to different concentrations of ammonia.

**METHODS:** In this study, Nile tilapia fingerlings were prepared and kept in the tub for adaptation, oxygenation, and feeding. A Range Finding test was performed on the fishes to find the ammonia killing range. Then, an acute toxicity test was performed on tilapia fish for 96 hours. For sub-lethal toxicity studies, 120 Nile tilapia fry were divided into 4 groups, including three groups based on different percentages of  $LC_{50}$  96 h (10, 20, and 30%  $LC_{50}$  96h) and a control group. The groups were exposed to ammonia for 14 days in an environment with constant temperature and pH control. Finally, histological examination and stress indices were performed on fish.

**RESULTS:** The results showed that a higher concentration of ammonia resulted in tissue damage, including discoloration and skin blackening, and behavioral changes, including swallowing air from the water surface, decreased appetite and decreased mobility which finally resulted in death in these animals ( $P \le 0.05$ ). The highest and lowest glucose levels were reported for the control group and 2.7 mg/L. In addition, stress indices were significantly increased in experimental groups ( $P \le 0.05$ ).

**CONCLUSIONS:** The data obtained from this study showed that increasing the amount of ammonia can lead to irreversible tissue damage to the structure of the skin and other tissues. In addition, the increase in oxidative stress in the Nile tilapia was due to the increase in ammonia, which led to lesions in these fish.

KEYWORDS: Ammonia, Histomorphometry, Nile tilapia, Skin, Stress

#### Correspondence

Hassan Morovvati, Department of Basic Sciences, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran. Tel: +98 (21) 61117117, Fax: +98 (21) 66933222, Email: hmorovvati@ut.ac.ir

Received: 2022-01-17

Accepted: 2022-03-16

Copyright © 2022. This is an open-access article distributed under the terms of the Creative Commons Attribution- 4.0 International License which permits Share, copy and redistribution of the material in any medium or format or adapt, remix, transform, and build upon the material for any purpose, even commercially.

#### How to Cite This Article

Rahmati, M., Morovvati, H., Abdi, R. (2022). Histomorphometric Analysis of Skin and Stress Indices of Nile Tilapia (*Oreo-chromis niloticus*) Exposed to Different Concentrations of Ammonia. *Iranian Journal of Veterinary Medicine*, 16(3), 288-297.

## Introduction

Fish is one of the most important sources of the human food chain. Due to climate change and the environmental effects of fishing in the ocean, fish farming is a priority for human societies (Adeli et al., 2011). Ammonia poisoning is one of the leading causes of fish mortality on farms. Ammonia with the chemical formula NH<sub>3</sub> is a combination of nitrogen and hydrogen. Ammonia is a stable binary hydride, and its simplest hydride is a colorless gas with a distinct pungent odor (Carlozzi and Padovani, 2016). Ammonia is a common nitrogenous waste, especially among aquatic organisms, and by acting as a precursor and in fertilizers, it significantly contributes to the nutritional needs of terrestrial organisms. Ammonia, directly or indirectly, is also a component of many pharmaceutical products and is used in many commercial cleaning products. This substance mainly accumulates by moving air and water downwards (Ogbonna and Chinomso, 2010). Because ammonia is one of the most significant excreted substances in aquatic environments produced by protein digestion, its toxicity has been studied extensively (Basir et al., 2012; Suliman et al., 2021).

Studies have shown that ammonia damages the skin, gills, and other tissues, leading to a variety of symptoms that may include respiratory problems such as rapid gill breathing, shortness of breath, swallowing air from the water surface, irregular swimming, and sudden darting movement, bruising of the skin (bleeding) and excessive production of mucus (dark spots on the skin), lethargy, loss of appetite, fish lying in bed, red streaks on the fins, body, and red or purple gills. Once the injury continues with ammonia poisoning, streaks or bloodstains appear on the body and fins. Internal damage also occurs to the brain, organs, and central nervous system. Bleeding occurs on the inner and outer surfaces of the fish and eventually leads to its death. Ammonia toxicity increases with acidity and water temperature (Morovvati et al., 2017; Dastan et al., 2017). Ammonia levels are usually controlled by biological filters that contain bacteria that break down ammonia into less toxic substances. Here, excreted ammonia is broken down by bacteria into nitrite and then nitrate (Jumaa et al., 2018). In addition, ammonia poisoning can be prevented mainly by routinely monitoring its levels using assay kits and avoiding high density of fish aqua culturing and overfeeding. Dangerous ammonia levels must be dealt with quickly, for example, by changing large amounts of water or using an ammonia-removing agent such as zeolite and new solutions (Morovvati *et al.*, 2017).

Following these quick steps makes more time until the underlying factor is identified (Jumaa et al., 2018). In the inland waters of Iran, there are 174 species of fish, some of which have economic value. Nile tilapia was first introduced to Iran from Indonesia and now forms an essential part of the fish farming basket in Iran (Azari et al., 2020; Khanjani and Sharifinia, 2021). The appearance of tilapia is very similar to that of sunfish or crappie. Still, its disconnected lateral line can easily be identified, which is characteristic of the cichlid family. Their body shape is compressed on the side, and they have long dorsal fins. The front of their dorsal fins is full of sharp bumps. These razors are visible on the ends of the anal fins and the anterior fins of the fish. Wide vertical lines are usually seen on both sides of the fingerlings and sometimes the body of the adult fish (Amin et al., 2019; El Asely et al., 2020). This species is an economically valuable fish for breeders. They reproduce easily, using a wide range of natural and artificial foods (Ibrahim et al., 2020). They cope with low water quality and grow rapidly in warm water. In addition to the low initial cost, these features make tilapia the most widely grown fish in the tropics and subtropics. This study aimed to study the effect of toxicity of different concentrations of ammonia on the skin and stress indices of the Nile tilapia.

## Materials and Methods

### **Fish Farming and Adaptation**

In this study, 120 juvenile Nile tilapia fishes  $(35\pm1)$  g) were transferred to the laboratory to adapt to environmental conditions. The prepared fish was kept in a 500-liter tub for 14 days. During this period, fish were fed commercial food (Bio mar, France daily for 2% of the fish's body weight by a plate under proper aeration. During the experiment, the average temperature of breeding water was  $27\pm1^{\circ}$ C, the oxygen concentration was  $6.2 \pm 1$  mg/L, and water hardness was  $269 \pm 3$  mg/L.

#### **Acute Toxicity Test**

In order to find the effect of acute ammonia toxicity, a range-finding test was performed (Adeli et al., 2011) by stationary method and according to standard instructions for 96 hours. Ammonia (Merck, Germany) was prepared as an ammonium chloride solution. The feeding of fish was stopped 24 hours before the acute toxicity test. All effective physicochemical parameters of water (pH, dissolved oxygen, and temperature) were monitored daily. The final test for acute ammonia toxicity was completed after determining the lethal range. For this purpose, 3 treatments with control groups of skin and tissue specimens were evaluated. The fish in each treatment were placed in a pre-aerated 15-liter aquarium. The dead fish were collected from the aquarium environment as soon as they were observed, and the number of losses was calculated and recorded at 24 hours intervals. After the test, the data obtained for the acute toxicity test were analyzed by Probit Analysis with a 95% confidence level.

#### **Sub-lethal Toxicity Test**

To evaluate the lethal toxicity of 120 Nile tilapia fingerlings, they were divided into 4 groups, three of which were selected based on different percentages of LC50 (10, 20, and 30% LC50 96 h) and placed with a control group. The fish were exposed to ammonia for two weeks in a temperature and pH-controlled environment (Campos et al., 2015; Pinto et al., 2016). The temperature and pH of the breeding water were monitored at 3 hours intervals. Therefore, 120 fingerlings were randomly distributed in 100-liter aquariums and subjected to a semi-static toxicity test (daily change of 20% of aquarium water). One hour before changing the water, feeding was done for 2% of the fish's body weight. The remaining food was removed from the aquariums by replacing water. To keep the ammonia concentration constant, an equal amount of water with ammonia was added to the aquarium, equal to the amount of water pouring out of the aquarium.

#### **Histological Examination**

Tissue sampling was performed randomly on the day of the experiment and 14 days after exposure. At

the end of the experimental period, the fish were randomly selected from each group and anesthetized using 0.5 g/L of clove powder. Then, sampling of the skin tissue of the head and trunk of the fish was performed using a tissue passage device (Automated tissue processors, Leica ASP300 S, Germany). Specimens were subjected to 10% formalin buffer, fixed, and transferred to 70% alcohol overnight (Mohamed et al., 2021; Mohamed et al., 2020). Tissue dehydration was performed using serial ethanol concentrations of 70, 80, 90, and 100%, and then the tissues were exposed to xylene. The tissues were then molded and paraffinized with paraffin at a melting point of 56-58°C on the tissue embedding console system. Section of 4 to 6 microns was prepared from the molds obtained using a microtome machine, and after placing on a slide, they were placed in an oven at 60°C for half an hour to remove excess paraffin from the tissue. The tissues were deparaffinized in xylene, rehydrated, and stained using hematoxylin and eosin solutions to stain the specimens. The slides were examined by a light microscope attached to a Dinolit lens as well as a computer system equipped with Dinocapture software. For histomorphometric valuation, five microscopic grounds per section and five sections per fish were exhausted.

## Results

#### **Survival Rate**

The study of survival rate in this study showed that no losses were observed during the adaptation period in different treatments under sublethal ammonia levels (0.9, 1.8, and 2.7 mg / l) and control during 14 days.

#### Histological Examination of Nile Tilapia Skin

The results of microscopic studies of fish skin exposed to different concentrations of ammonia compared with those studied in the control group showed changes such as an increase in the number of mucosal and caveat cells, an increase in the number of epithelial cells, and also an increase in the thickness of the epidermis, which regulated histological results. No specific cell lesion was seen. Overall, the severity of these complications increased significantly from low to high concentration treatment ( $P \le 0.05$ ) (Figures 1 and 2).



**Figure 1.** Tissue micrograph of Nile tilapia skins in experimental groups (H&E, x40) A) Epidermis (Ep), dermis (De), caveat cell (Cc), hyperplasia (H) in control groups, B) Exposure to ammonia 0.9 mg/L., C) Exposure to ammonia 1.8 mg/L., D) Exposure to ammonia 2.7 mg/L during 14 days.



**Figure 2.** Tissue micrograph of the middle skin of the Nile tilapia trunk of experimental groups (H&E, x40). Epidermis (Ep), Dermis (De), Caveat cell (Cc), Hyperplasia (H), Melanin pigment (Mp), Scales (Sc), Mucosal cell (Gc), Collagen filament (Co), Mucosal cell proliferation (Thick arrows) and proliferation of caveat cells (thin arrows) in control groups, A) Exposure to 0.9 mg/L ammonia, B) Exposure to ammonia 1.8 mg/L, C) Exposure to 2.7 mg/L ammonia during two weeks.

#### Level of Stress Indices during Adaptation Period

The results of measuring glucose and cortisol levels during the adaptation period in different treatments are shown in the following diagrams. Results showed a significant increase in cortisol and glucose levels in almost all treatments compared to the control group ( $P \le 0.05$ ). Also, after exposure of fish to ammonia, the maximum and minimum glucose levels were reported as 27.19  $\pm$ 1.52 mg/dL and 50.35  $\pm$ 3.19 mg/dL for the control specimens, respectively.



**Figure 3.** Figure A shows the increase in the thickness of the caudal stalk epidermis, Figure B shows the increase in the mucosal cells of the head specimens, Figure C shows the increase in the thickness of the epidermis of the middle body skin specimens, Figure D shows the increase in the thickness of the epidermis of the head specimen, Figure E shows the increase of the middle body epidermal thickness, and Figure F shows the increase of the mucosal cells of the caudal stalk epidermis, (Different symptoms indicate a significant difference in the level of P<0.05).



**Figure 4.** Measurement of cortisol (left chart) and glucose (right chart) levels in ammonia-treated groups 0.9 mg/L (A), 1.8 mg/L (B), and 2/7 mg/L (C) compared to control during two weeks were shown (Different symptoms indicate a significant difference in level of P<0.05).

## Discussion

Today environmental problems, inefficiency, and the high cost of fishing in the sea have led to increased fish farming as a suitable alternative. Providing inexpensive and valuable food is one of the most important goals of the food industry worldwide (Wanka et al., 2018). In recent years, Nile tilapia has become one of the most common fish for breeding and consumption worldwide, ranking second after carp species (Abdi et al., 2011). Asia is the top tilapia producer and accounts for more than 70% of global tilapia production. Many diseases in aquatic life cause a loss of capital and energy and increase the risk of fish farming (Føre et al., 2018). Ammonia toxin is one of the most important chemicals that destroy aquatic tissues, which is produced due to protein digestion by these organisms and can lead to increased mortality among them (Zuffo et al., 2021). Ammonia in aqueous media can be detected in both ionized and non-ionized forms. The non-ionized type of which has higher toxicity due to its high permeability through the epithelium of aquatic organisms (Abdel-Tawwab et al., 2020; Nogueira et al., 2018). So far, the effects of different concentrations of water-soluble ammonia on the Nile tilapia have not been studied, so this study aimed to investigate the effect of this toxin on the skin histomorphology and stress indices of this fish. The present study's data showed that increasing ammonia concentration in the experimental group results in appetite and mobility decrease, skin discoloration and blackening, significant histomorphologic responses, and an increase in stress indices in aquatic animals (Deswati et al., 2018). In this study, poisoning symptoms appeared in high concentrations of ammonia and the last days, followed by nervous behavioral symptoms. On the skin's outer surface, pallor and bleeding mucus accumulation were evident on the body's surface. The skin tissue of the fish changed, such as an increase in the number of mucus cells and caveat cells, which led to an increase in the thickness of the epidermis. In general, the severity of these factors was higher from low concentration treatment to high concentration treatment and had a statistically significant difference compared to control. These experiments also showed that increasing ammonia caused a significant increase in cortisol

and glucose in most treatments compared to the control group. El Shafi *et al.*, in a study, examined the chronic effects of ammonia on the Nile tilapia. This experiment was performed for 75 days using fish with an average body weight of 20 grams. Five nominal concentrations of total ammonia nitrogen (control, 2.5, 5, 7.5, and 10 mg) were generated as treatment groups. The data showed that increasing the ammonia concentration increased the mortality rate.

This study concluded that for the cultivation of Nile tilapia in duckweed-fed ponds, due to the negative effect of ammonia on growth performance, its level should be maintained below 0.1 mg/L (El-Shafai et al., 2014). In another study, Aysel et al. examined the lethal effects of ammonia on Nile tilapia. Their study aimed to determine the mean lethal concentration (LC50) of ionized ammonia for larvae and juveniles of Nile tilapia. The mean 48-hour LC50 values for larvae and juveniles were 1.009±0.02 mg/L and 7.40±0.01 mg /L, respectively. Larvae exposed to different ammonia concentrations are motivated very quickly to lose their balance in the water and swim sideways. In fingerlings, increased movements, ventilation, seizures, spiral swimming, attempts to swallow air from the water surface, increased mucous secretions in the gills and body surface, bleeding in the gills, and skin darkness were observed. Histological studies showed hyperplasia and lamella fusion in gills (Benli and Köksal, 2005). A study by Benli et al. also confirmed the negative effects of high ammonia concentrations on the liver, gills, and kidneys of Nile tilapia (Benli et al., 2008). The findings of the present study in the field of behavioral changes as well as the toxic effects of ammonia were following the findings of the mentioned studies. In addition, the study results by Jian et al. confirmed the effects of ammonia on the incidence of behavioral responses in Nile tilapia (Jianyu et al., 2005), which is in agreement with the present study. Another study by Yilmaz et al. (2020) confirms the data of the Magouz (Magouz et al., 2021) study and the present study on the occurrence of behavioral changes in the presence of high ammonia levels in the water. Mercante et al. (2018) also confirmed the increase in tissue problems due to the

increase in the duration of exposure of fish to aquatic ammonia. They showed that this exposure could increase cortisol and oxidative stress leading to death in studied aquatic animals. The results of Yousefi *et al.* (2020) research showed that after 12 hours of exposure to ammonia, the fish changed the initial treatments of stress factors. Cortisol increased in the first two treatments and then decreased in the following three treatments. But it was still higher than control cortisol in all treatments. The glucose levels reached their lowest level in the first treatment and showed a significant increase in other treatments (Yousefi *et al.*, 2020). In the present study, increasing the ammonia concentration led to a significant increase in stress factors.

#### Conclusion

The present study's data indicate an increase in the negative effects of ammonia on the skin and the level of stress indices in Nile tilapia. These results emphasize the monitoring of ammonia levels in the breeding environment of these fish and prove that increasing the concentration of ammonia can lead to increased behavioral responses by these organisms in addition to causing tissue problems.

### Acknowledgments

The authors wish to express their appreciation to everyone that assists us in this study.

## **Conflict of Interest**

The authors declared that there is no conflict of interest.

## References

- Abdel-Tawwab, M., Monier, M. N., Abdelrhman, A. M., & Dawood, M. A. (2020). Effect of dietary multi-stimulants blend supplementation on performance, digestive enzymes, and antioxidants biomarkers of common carp, Cyprinus carpio L. and its resistance to ammonia toxicity. *Aquaculture*, *528*, 735529. [DOI:10.1016/j.aquaculture.2020.735529]
- Abdi, R., Pourkhadje, M. R., Zolgharnein, H., Hosseinzadeh Sahafi, H., & Morovvati, H. (2011). Effect of salinity on mitochondria of chloride cells in gill of juvenile's grouper (Epinephelus coioides). *Journal of Animal Environment*, 2(4), 37-42.
- Adeli, A., Hasangholipour, T., Hossaini, A., Salehi, H., & Shabanpour, B. (2011). Status of fish Consumption per capita of Tehran citizens. *Iranian Journal of Fisheries Sciences*, 10(4), 546-556.
- Amin, A., El Asely, A., Abd El-Naby, A. S., Samir, F., El-Ashram, A., Sudhakaran, R., & Dawood, M. A. (2019).
  Growth performance, intestinal histomorphology and growth-related gene expression in response to dietary Ziziphus mauritiana in Nile tilapia (Oreochromis niloticus). *Aquaculture*, *512*, 734301.
  [DOI:10.1016/j.aquaculture.2019.734301]

- Azari, A., Anvar, S. A. A., Ahari, H., Sharifan, A., & Motallebi Moghanjoghi, A. A. (2020). Study on Nanosilver-TiO2 photocatalytic nanocomposite coating with extrusion technique for increasing shelf life of Nile Tilapia (Oreochromis niloticus). *Iranian Journal* of Fisheries Sciences, 19(5), 2618-2633.
- Basir, Z., Morovvati, H., Khaksari, M., Mmesbah, M., & Abdi, R. (2012). Histomorphology and Histometric Study of the Head's Skin in (Barbus grypus). *Journal* of Cell and Tissue, 3(1): 73-81.
- Benli, A. Ç. K., & Köksal, G. (2005). The acute toxicity of ammonia on tilapia (Oreochromis niloticus L.) larvae and fingerlings. *Turkish Journal of Veterinary and Animal Sciences*, 29(2), 339-344.
- Benli, A. Ç. K., Köksal, G., & Özkul, A. (2008). Sublethal ammonia exposure of Nile tilapia (Oreochromis niloticus L.): Effects on gill, liver and kidney histology. *Chemosphere*, 72(9), 1355-1358.
  [DOI:10.1016/j.chemosphere.2008.04.037] [PMID]
- Campos, B. R. D., Furtado, P. S., & Poersch, L. H. D. S. (2015). The chronic toxicity of ammonia, nitrite and nitrate on juvenile Farfantepenaeus brasiliensis (Crustacea: Decapoda). *Boletim do Instituto de Pesca*, 41(2): 261-269.

- Carlozzi, P., & Padovani, G. (2016). The aquatic fern Azolla as a natural plant-factory for ammonia removal from fish-breeding fresh wastewater. *Environmental Science and Pollution Research*, 23(9), 8749-8755.
   [DOI:10.1007/s11356-016-6120-8] [PMID]
- Dastan, V., Abdi, R., Movahedinia, A. A., & Aliabadi, S. (2017). Study of gill and kidney tissue changes in Tenualosa ilisha during migration from sea to the Karun and Bahmanshir rivers. *Iranian Scientific Fisheries Journal*, 25(4): 53-62.
- Febriani, N., Pardi, H., Yusuf, Y., & Suyani, H. (2018).
  Applications of aquaponics on Pakcoy (Brassica Rapa L) and Nila Fish (Oreochromis Niloticus) to the concentration of ammonia, nitrite, and nitrate. *Oriental Journal of Chemistry*, 34(5), 2447.
  [DOI:10.13005/ojc/340529]
- El Asely, A. M., Reda, R. M., Salah, A. S., Mahmoud, M. A., & Dawood, M. A. (2020). Overall performances of Nile tilapia (Oreochromis niloticus) associated with using vegetable oil sources under suboptimal temperature. *Aquaculture Nutrition*, 26(4), 1154-1163. [DOI:10.1111/anu.13072]
- El-Shafai, S. A., El-Gohary, F. A., Nasr, F. A., van der Steen, N. P., & Gijzen, H. J. (2004). Chronic ammonia toxicity to duckweed-fed tilapia (Oreochromis niloticus). *Aquaculture*, 232(1-4), 117-127.
  [DOI:10.1016/S0044-8486(03)00516-7]
- Føre, M., Frank, K., Norton, T., Svendsen, E., Alfredsen, J. A., Dempster, T., ... & Berckmans, D. (2018). Precision fish farming: A new framework to improve production in aquaculture. *Biosystems Engineering*, *173*, 176-193. [DOI:10.1016/j.biosystemseng.2017.10.014]
- Ibrahim, R. E., Ahmed, S. A., Amer, S. A., Al-Gabri, N. A., Ahmed, A. I., Abdel-Warith, A. W. A., ... & Metwally, A. E. (2020). Influence of vitamin C feed supplementation on the growth, antioxidant activity, immune status, tissue histomorphology, and disease resistance in Nile tilapia, Oreochromis niloticus. *Aquaculture Reports*, 18, 100545. [DOI:10.1016/j.aqrep.2020.100545]
- Jian-yu, X., Xiang-wen, M., Ying, L., & Shao-rong, C. (2005). Behavioral response of tilapia (Oreochromis niloticus) to acute ammonia stress monitored by computer vision. *Journal of Zhejiang university science B*, 6(8), 812-816. [DOI:10.1631/jzus.2005.B0812] [PMID] [PMCID]
- Jumaa, Q. S., Al-Saeedy, K. A. H., & Obid, W. S. (2018). The toxic effects of ammonia on growth rate and some

blood parameters in common carp (Cyprinus carpio L.). *Al-Anbar Journal of Veterinary Sciences*, *11*(1). [DOI:10.37940/AJVS.2018.11.1.13]

- Khanjani, M. H., & Sharifinia, M. (2021). Production of Nile tilapia Oreochromis niloticus reared in a limited water exchange system: The effect of different light levels. *Aquaculture*, 542, 736912. [DOI:10.1016/j.aquaculture.2021.736912]
- Magouz, F. I., Mahmoud, S. A., El-Morsy, R. A., Paray, B. A., Soliman, A. A., Zaineldin, A. I., & Dawood, M. A. (2021). Dietary menthol essential oil enhanced the growth performance, digestive enzyme activity, immune-related genes, and resistance against acute ammonia exposure in Nile tilapia (Oreochromis niloticus). *Aquaculture*, 530, 735944.
  [DOI:10.1016/j.aquaculture.2020.735944]
- Mercante, C. T., David, G. S., Rodrigues, C. J., do Carmo, C. F., & da Silva, R. J. (2018). Potential toxic effect of ammonia in reservoirs with Tilapia culture in cages. *International Journal of Fisheries and Aquatic Studies*, 6(5), 256-261.
- Mohamed, M., Abdi, R., Ronagh, M. T., Abadi, S. A., & Basir, Z. (2021). Comparative histomorphology of epidermis of head and caudal peduncle in Otolithes ruber, Huso huso and Pangasius hypophthalmus fish. *Iranian Journal of Aquatic Animal Health*, 7(1), 10-20. [DOI:10.52547/ijaah.7.1.10]
- Mohamed, M., Abdi, R., Ronagh, M. T., Salari Ali Abadi, M. A., & Basir, Z. (2020). Comparative histomorphometry of dorsal, ventral and lateral skin in macroscopy, microscopy and free scale fish. *Iranian Veterinary Journal*, 16(2), 47-53.
- Morovvati, H., Abdi, R., & Shamsi, M. M. (2017). Effect of different salinity concentration on gill of benni Barbus sharpeyi. *Journal of Veterinary Research*, 72(2).
- Morovvati, H., Zolgharnein, H., Noori Moghahi, M. H., Abdi, R., & Ghazilou, A. (2012). Alterations to chloride cells of the secondary lamella and gill branches of spotted scat (Scatophagus argus L.) in different salinities. *Journal of Veterinary Research*, 67(2), 109-117.
- Nogueira, S. M. S., Souza, J., Maia, H. D., Saboya, J. P. S., & Farias, W. R. L. (2018). Use of Spirulina platensis in treatment of fish farming wastewater. *Revista Ciência Agronômica*, 49, 599-606. [DOI:10.5935/1806-6690.20180068]
- Ogbonna, J., & Chinomso, A. (2010). Determination of the concentration of ammonia that could have lethal effect on fish pond. *Journal of Engineering and Applied Sciences (Asian Research Publishing Network)*, *5*, 1-5.

- Pinto, D. S. B., Maltez, L. C., Stringhetta, G. R., Pellegrin, L., Nitz, L. F., Figueiredo, M. R. C., & Garcia, L. D. O. (2016). Ammonia and nitrite acute toxicity in juvenile piavuçu Leporinus macrocephalus (Actinopterygii, Anostomidae). *Pan-American Journal of Aquatic Sciences*, 11(4), 292-300.
- Suliman, E. A. M., Osman, H. A., & Al-Deghayem, W. A. A. (1970). Histopathological changes induced by ectoparasites on gills and skin of Oreochromis niloticus (Burchell 1822) in fish ponds. *Journal of Applied Biology and Biotechnology*, 9(1), 6-4.
- Wanka, K. M., Damerau, T., Costas, B., Krueger, A., Schulz, C., & Wuertz, S. (2018). Isolation and characterization of native probiotics for fish farming. *Bmc Microbiology*, 18(1), 1-13. [DOI:10.1186/s12866-018-1260-2] [PMID] [PMCID]

- Yilmaz, E. (2020). Effect of dietary carob (Ceratonia siliqua) syrup on blood parameters, gene expression responses and ammonia resistance in tilapia (Oreochromis niloticus). *Aquaculture Research*, 51(5), 1903-1912. [DOI:10.1111/are.14540]
- Yousefi, M., Vatnikov, Y. A., Kulikov, E. V., Plushikov, V. G., Drukovsky, S. G., Hoseinifar, S. H., & Van Doan, H. (2020). The protective effects of dietary garlic on common carp (Cyprinus carpio) exposed to ambient ammonia toxicity. *Aquaculture*, *526*, 735400. [DOI:10.1016/j.aquaculture.2020.735400]
- Zuffo, T. I., Durigon, E. G., Morselli, M. B., Picoli, F., Folmann, S., Kinas, J. F., ... & Lopes, D. L. D. A. (2021). Lethal temperature and toxicity of ammonia in juveniles of Curimbatá (Prochilodus lineatus). *Aquaculture*, 545, 737138. [DOI:10.1016/j.aquaculture.2021.737138].

Abstracts in Persian Language

مجله طب دامی ایران، ۱۴۰۱، دوره ۱۶، شماره ۳، ۲۸۸–۲۹۷

# تجزیه و تحلیل هیستومورفومتریک پوست و شاخصهای استرس ماهی تیلاپیا نیل (*Oreochromis niloticus*) در معرض غلظتهای مختلف آمونیاک

معصومه رحمتی 🔟، حسن مروتی'، رحیم عبدی'

<sup>ا ت</sup>کروه علوم پایه، دانشکده دامپزشکی، دانشگاه تهران، تهران، ایران <sup>ام</sup>روه زیستشناسی دریا، دانشکده علوم دریایی، دانشگاه علوم و فنون دریایی خرمشهر، خرمشهر، ایران

(دریافت مقاله: ۲۷ دی ماه ۱۴۰۰، پذیرش نهایی: ۲۵ اسفند ۱۴۰۰)

**زمینه مطالعه**: آبزی پروری به صورت فشرده همواره با غلظت بالای آلایندههای نیتروژن همراه است، که این امر میتواند منجر به افزایش خسارت اقتصادی و اتلاف انرژی و افزایش مرگ و میر ماهیها شود.

هدف: این مطالعه با هدف بررسی هیستومورفومتری پوست و شاخصهای استرس ماهی تیلاپیای نیل طی مواجهه با غلظتهای مختلف آمونیاک انجام شد.

روش کار: در این مطالعه ماهیان جوان تیلاپیا نیل تهیه و برای سازگاری، هوادهی و تغذیه در وان نگهداری شدند. آزمایش محدوده یابی بر روی ماهیها برای یافتن محدوده کشنده آمونیاک انجام شد. سپس آزمایش سمیت حاد بر روی ماهی تیلاپیا به مدت ۹۶ ساعت انجام گرفت. برای مطالعات سمیت تحت کشنده، ۱۲۰ قطعه بچه ماهی تیلاپیا نیل به ۴ گروه شامل: سه گروه بر اساس درصدهای مختلف 96h الاری الار، ۲۰ و ۳۰ درصد 96h و LC50 و یک کشنده، ۱۲۰ قطعه بچه ماهی تیلاپیا نیل به ۴ گروه شامل: سه گروه بر اساس درصدهای مختلف 96h الاری الار، ۲۰ و ۳۰ درصد 96h و یک الاری و یک گروه کنترل تقسیم شدند. گروهها به مدت ۱۴ مواند در نهایت مطالعات بافتی و الاری کنترل شده در معرض آمونیاک قرار گرفتند. در نهایت مطالعات بافتی و فاکتورهای استرس بر روی ماهیان انجام گرفت. در نهایت مطالعات بافتی و فاکتورهای استرس بر روی ماهی تعلی معانده مواندی الالالات بافتی و و کنترل شده در معرض آمونیاک قرار گرفتند. در نهایت مطالعات بافتی و فاکتورهای استرس بر روی ماهیان انجام گرفت.

نتایج: نتایج نشان داد که غلظت بالاتر آمونیاک منجر به آسیب بافتی از جمله تغییر رنگ و سیاه شدن پوست و تغییرات رفتاری از جمله بلع هوا از سطح آب، کاهش اشتها،کاهش تحرک و در نهایت منجر به مرگ ماهی شد (۵۰/۰≤P ). بالاترین و کمترین میزان گلوکز به ترتیب برای گروه کنترل و تیمار ۲/۷ میلیگرم در گزارش شده که پس از مواجهه با آمونیاک بهطور معنیداری افزایش یافت (۵۰/۰≤P ). همچنین شاخصهای استرس در گروههای آزمایشی بهویژه در مقادیر بالا بهطور معنی داری افزایش یافت (۵۰/۰≤P).

**نتیجهگیری نهایی:** دادههای بهدستآمده از این مطالعه نشان داد که افزایش میزان آمونیاک میتواند منجر به آسیب غیرقابل برگشت بافتی به ساختار پوست و سایر بافتها شود. علاوه بر این، افزایش استرس در تیلاپیا نیل به دلیل افزایش آمونیاک که منجر به ضایعاتی در این ماهیها شد.

واژههای کلیدی: آمونیاک، هیستومورفومتری، تیلاپیا نیل، پوست، استرس

نویسندهٔ مسئول: حسن مروتی، گروه علوم پایه، دانشکده دامپزشکی، دانشگاه تهران، تهران، ایران ایمیل.<u>hmorovvati@ut.ac.ir</u>